

Prior hydration of *Brassica tournefortii* seeds reduces the stimulatory effect of karrikinolide on germination and increases seed sensitivity to abscisic acid

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- **Background and Aims** The smoke-derived compound karrikinolide (KAR₁) shows significant potential as a trigger for the synchronous germination of seeds in a variety of plant-management contexts, from weed seeds in paddocks, to native seeds when restoring degraded lands. Understanding how KAR₁ interacts with seed physiology is a necessary precursor to the development of the compound as an efficient and effective management tool. This study tested the ability of KAR₁ to stimulate germination of seeds of the global agronomic weed *Brassica tournefortii*, at different hydration states, to gain insight into how the timing of KAR₁ applications in the field should be managed relative to rain events.
- **Methods** Seeds of *B. tournefortii* were brought to five different hydration states [equilibrated at 15 % relative humidity (RH), 47 % RH, 96 % RH, fully imbibed, or re-dried to 15 % RH following maximum imbibition] then exposed to 1 nM or 1 µM KAR₁ for one of five durations (3 min, 1 h, 24 h, 14 d or no exposure).
- **Key Results** Dry seeds with no history of imbibition were the most sensitive to KAR₁; sensitivity was lower in seeds that were fully imbibed or fully imbibed then re-dried. In addition, reduced sensitivity to KAR₁ was associated with an increased sensitivity to exogenously applied abscisic acid (ABA).
- **Conclusions** Seed water content and history of imbibition were found to significantly influence whether seeds germinate in response to KAR₁. To optimize the germination response of seeds, KAR₁ should be applied to dry seeds, when sensitivity to ABA is minimized.

Key words: Karrikinolide, karrikins, butenolide, smoke, germination stimulant, seed water content, abscisic acid, ABA, gibberellin, weed, *Brassica tournefortii*.

INTRODUCTION

When seeds are dispersed from the parent plant, they can be dormant and unable to germinate despite exposure to environmental conditions that might otherwise be suitable for germination. Seeds that are physiologically dormant will not germinate until dormancy is alleviated by processes such as dry after-ripening, wet–dry cycles, or cold- or warm-stratification (Merritt *et al.*, 2007; Hoyle *et al.*, 2008). Such dormancy alleviation processes are mediated by seed water content and the temperature of the environment (Merritt *et al.*, 2007). Seeds with a permeable seed coat will equilibrate with ambient moisture conditions (Wuest, 2007), whether seeds are on the surface of the soil, buried or retained serotiously on the parent plant.

As an alternative to natural alleviation of dormancy, seeds can be stimulated to germinate by chemical cues, such as nitrates (Finch-Savage *et al.*, 2007), strigolactones (Goldwasser *et al.*, 2008) and a suite of chemicals present in smoke (Chiwocha *et al.*, 2009). One such seed germination stimulant identified from smoke is karrikinolide (KAR₁ – a butenolide) (Flematti *et al.*, 2004). Karrikinolide triggers the

germination of numerous species from fire-prone (Dixon *et al.*, 2009), non-fire-prone (Merritt *et al.*, 2007) and agricultural environments (Daws *et al.*, 2007; Stevens *et al.*, 2007), highlighting its potential for use in a range of plant-management contexts. Applying KAR₁ to mined and degraded land could assist with regenerating flora from broadcast seed or seeds within replaced topsoil (Commander *et al.*, 2008, 2009). In a commercial-seed context, KAR₁ may be used as a priming agent to enhance the subsequent germination characteristics of crop seeds (Jain and Van Staden, 2007). Furthermore, in agricultural systems, weeds could be triggered to germinate on demand, maximizing the efficiency of herbicide applications, and reducing the burden of pesticides in the environment (Davis *et al.*, 2005; Stevens *et al.*, 2007). To realize such valuable applications for KAR₁ and ensure its efficacy, it is necessary to explore its effect under a variety of environmental conditions. In particular, studies that optimize the timing of application of KAR₁ relative to when seeds are most receptive would contribute to the efficient use of KAR₁ in agricultural and restoration environments.

Environmental conditions may alter the receptivity of seeds to smoke signals (Roche *et al.*, 1998; Baker *et al.*, 2005).

For example, smoke has been shown to be more effective at eliciting germination when applied during the dry autumn of a Mediterranean climate than when applied during the wetter months of winter and spring (Roche *et al.*, 1998). These observations imply that there may be an interaction between the hydration state of seeds and the efficacy of smoke-derived chemicals such as KAR₁. To date, studies of KAR₁ and related smoke-extracts have not considered how the hydration state of seeds interacts with stimulatory chemicals to influence germination outcomes. However, an influence of hydration state on germination is likely, because seed water content affects both the physical and the physiological characteristics of seeds.

Physical studies of imbibing seeds indicate that the rate and volume of solution uptake is greatly influenced by seed water potential (Bradford, 1995). Drier seeds have more negative matric water potentials and absorb water more quickly than moister seeds with less negative water potentials (Wolk *et al.*, 1989). In contrast, uptake of solutions by fully imbibed seeds is driven by differences in osmotic potential between the seed and the surrounding solution (Roberts and Ellis, 1989). Thus, the water content of seeds may be an important factor affecting the uptake of aqueous stimulants and subsequent germination response.

Whilst the hydration state of seeds may impart physical effects on the response of seeds to germination stimulants, it may also influence the response physiologically. The water content of seeds directly affects molecular mobility within cells, and consequently the types and rates of reactions that occur (Walters *et al.*, 2005). Moderate seed hydration [e.g. -100 to -10 MPa, 40–92 % relative humidity (RH)] favours degradative reactions, whilst lower seed water contents (10–20 % RH) can induce a state of relative molecular paralysis that optimizes seed longevity (Walters *et al.*, 2005). Imbibition to seed hydration states above approx. -10 MPa provides sufficient water for reparative reactions to occur (Bewley, 1997; Walters *et al.*, 2005). Indeed, in priming, seeds can be fully imbibed then re-dried to improve subsequent germination percentages and rates (Gurusinghe *et al.*, 2002; Demir *et al.*, 2009). The increased physiological activity associated with imbibition can alter the balance of compounds that regulate germination, or the sensitivity of seeds to these compounds (Bu *et al.*, 2009). Pertinently for seed dormancy, imbibition permits synthesis or breakdown of plant growth regulators such as abscisic acid (ABA; Grappin *et al.*, 2000; Ali-Rachedi *et al.*, 2004), which maintains dormancy and inhibits germination, and gibberellins (GA), which promote germination (Nelson *et al.*, 2009). These plant growth regulators may in turn interact antagonistically or synergistically with germination stimulants such as KAR₁, although the nature and mechanisms of these interactions remain unclear (Nelson *et al.*, 2009).

Considering the many ways that the hydration state of seeds affects their physical and physiological characteristics, we reasoned that the hydration state of seeds will influence their germination response to KAR₁. More specifically, KAR₁ will stimulate more germination when applied to dry seeds than to seeds with higher water contents. In addition, based on the key roles of ABA and GA in maintaining dormancy and inducing seed germination, the interaction of these plant growth regulators with seed hydration state and sensitivity to KAR₁ was examined.

MATERIALS AND METHODS

Seed material

The annual weed *Brassica tournefortii* was chosen for this study as it has been shown to be highly responsive to KAR₁ (Stevens *et al.*, 2007). Seeds of *B. tournefortii* were collected from wild plants growing in semi-natural dune vegetation at City Beach, Perth, Western Australia (31°53'26''S, 115°45'21''E) in October, 2006. Following drying at 15 % RH and 15 °C for 4 weeks, seeds were sealed in a foil bag and stored at 5 °C until used in experiments.

Initial seed hydration states

Brassica tournefortii seeds were prepared according to one of five initial seed hydration treatments: 15 % RH, 47 % RH, 96 % RH, fully imbibed, or fully imbibed then dried to 15 % RH ('re-dried') prior to exposure to KAR₁. For all hydration treatments, multiple sets of 50 seeds were sealed in nylon mesh bags. Samples for the 15 % RH treatment were stored in a controlled-environment room at 15 % RH and 15 °C. Samples for the 47 and 96 % RH treatments were placed in sealed boxes (polycarbonate electrical enclosure boxes; 270 × 190 × 100 mm; NHP Fibox, Perth, Australia) and suspended above solutions of LiCl (330 g L⁻¹) and saturated KNO₃, respectively, to equilibrate for 4 weeks at 15 °C. For the fully imbibed treatment, samples were placed in tissue culture-grade deionized water (TC-H₂O) at room temperature (21 °C) for 18 h, using an automatic shaker (40 r.p.m.; Orbital Mixer, Ratek Instruments, Sydney, Australia) to gently agitate the water. Similarly, seeds in the re-dried treatment were allowed to imbibe for 18 h then returned to the 15 % RH environment (15 °C) to dry for 26 h.

KAR₁ sensitivity

For each hydration treatment (see above) three replicate samples were exposed to 200 mL of 1 nM or 1 μM KAR₁ (synthesized from pyromeconic acid according to Flematti *et al.*, 2005) for 0 min, 3 min, 1 h, 24 h or 14 d. Three samples for each combination of hydration treatment and KAR₁ exposure duration were placed in separate replicate containers of 1 nM or 1 μM KAR₁ and agitated on an automated shaker. Samples exposed to KAR₁ for ≤24 h were subsequently rinsed in 400 mL TC-H₂O and transferred to individual 90-mm Petri dishes containing 20 mL TC-H₂O for the remainder of the 24-h period, so that all samples were hydrated for the same period. Samples were then patted dry and transferred to 90-mm Petri dishes containing 1 % (w/v) agar for germination. For samples exposed to KAR₁ for 14 d, seeds were removed from the KAR₁ soaking solution after 24 h, patted dry and transferred directly to germination media containing either 1 nM or 1 μM KAR₁.

Germination testing

Petri dishes were wrapped in two layers of aluminium foil to exclude light and incubated at 20 °C (Thermoline, Brisbane, Queensland, Australia). Germination was scored once after 14 d, and seeds were scored as germinated if the radicle had

visibly protruded (>1 mm). Seedlings that germinated but which failed to develop normally (e.g. radicle appeared glassy) were not included in germination totals. Any seeds that did not germinate were inspected for obvious necrosis and squeezed with forceps to assess viability; seeds with firm white embryos were scored as viable.

Imbibition curves

Seed water uptake was monitored over a 24-h period for each of the initial hydration states (see above) to create imbibition curves. Due to the mucilaginous layer surrounding *B. tournefortii* seeds, which was degraded when patted dry prior to weighing samples, a destructive method of sampling was adopted. Thirty nylon mesh bags containing 50 seeds were prepared for each initial hydration state. The weight of seeds (at 15 % RH) in each bag, and the seed + bag weights were recorded prior to applying the hydration treatments. Ten bags of each hydration treatment were placed in each of three replicate solutions of TC-H₂O to imbibe. Periodically, one bag for each hydration treatment and replicate solution was removed from the solution, patted dry to remove excess water then weighed. For each sample, seed weight gain was calculated as (current weight – original weight at 15 % RH)/(original weight at 15 % RH) × 100 %.

Scanning electron microscopy

Scanning electron microscopy was used to compare the surface features of seeds prepared according to the re-dried treatment and seeds at 15 % RH with no history of imbibition; fully imbibed samples could not be imaged as only dry samples could be tested. Nine seeds of each treatment were gold-plated (Emitech K550X sputter-coater, Quorum Technologies Ltd, East Grinstead, UK) and viewed with a Jeol NeoScope JCM 5000 scanning electron microscope (10 kV acceleration voltage, 10⁻² Pa; Tokyo, Japan).

ABA and GA concentrations

Three samples of seeds (approx. 500 mg at 15 % RH, actual weight recorded) were pre-equilibrated to each of the five target hydration states in nylon mesh bags, as described above. Samples were ground to a fine powder in a mortar and pestle with liquid nitrogen and analysed for their ABA, GA₁, GA₃ and GA₄ content according to the methods of Nelson *et al.* (2009). Only ABA, GA₃ and GA₄ were detected in the samples.

ABA, GA₃ and GA₄ sensitivity

To investigate further the effect of seed hydration on seed responses to KAR₁, the sensitivity of seeds to exogenously applied ABA and GA₃ and GA₄ were assessed for the 15 % RH, fully imbibed and re-dried treatments. Germination of *B. tournefortii* was assessed in response to six concentrations of biologically active (+)-ABA (0, 0.5, 1, 5, 10 or 50 μM, Sigma-Aldrich Company, St Louis, MO, USA) in combination with 1 μM KAR₁. Six concentrations of GA₃ and GA₄ (Sigma-Aldrich) were also tested to assess the sensitivity of

seeds to GA. The GA₃ concentrations used were 0, 0.5, 1, 5, 10 and 100 μM, and the GA₄ concentrations were 0, 0.1, 0.5, 1, 5 and 10 μM. All ABA and GA media were prepared by adding stock solutions of the plant-growth regulators to 1 % (w/v) agar, after the agar was autoclaved (121 °C for 20 min) and allowed to cool to approx. 60 °C. Three replicates of 50 seeds for each hydration state were assigned to each treatment in the ABA and GA sensitivity experiments. Conditions for germination were consistent with the KAR₁ sensitivity experiment.

Seed water content assessment

For each experiment, all initial hydration treatments were tested for their water content and RH to verify that the treatments were consistent. Water content was assessed gravimetrically by weighing three samples of each treatment before and after drying them in an oven at 105 °C for 48 h. Seed water content was calculated on a dry-weight percentage basis: [(fresh weight – dry weight)/dry weight] × 100 (Table 1).

Statistical analyses

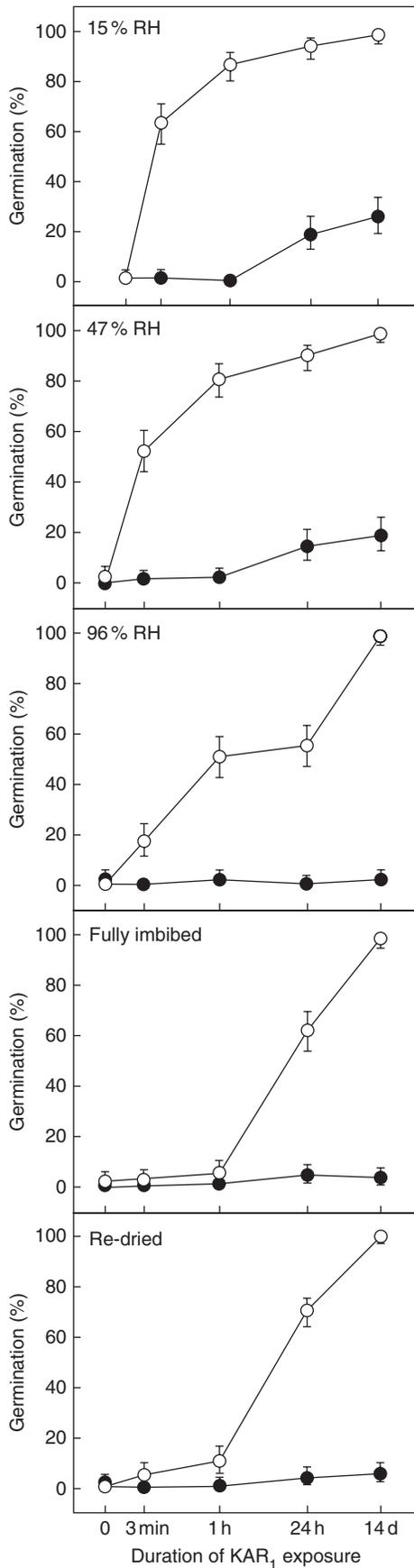
A generalized linear model with a logit link-function was fitted to germination data using step-wise regression (GenStat 10th Edition; McCullagh and Nelder, 1989). For the results of the KAR₁ exposure experiment, the terms included in the model were KAR₁ concentration, exposure time, seed hydration state, KAR₁ concentration × exposure time, and seed hydration state × exposure time (all terms, *P* < 0.001). For the analysis of the ABA sensitivity experiment, the terms included in the model were seed hydration state, concentration and seed hydration state × concentration (all terms, *P* < 0.001); the only term that had a significant effect on modelling the GA sensitivity data was GA concentration. To test for differences in the endogenous concentration of ABA, GA₃ and GA₄, a generalized linear model with a log-arithm link-function was fitted to the data. In all experiments, there was no bias based on replicate results. Comparisons of individual hydration states were conducted using parameter estimates from models including only the seed hydration state term. Data for the effect of hydration state on the germination response to KAR₁ (Fig. 1), ABA sensitivity (Fig. 5A) and GA sensitivity (Fig. 5B, C) were transformed via log(*x* + 1) to facilitate graphing, but non-transformed data were used in all statistical analyses.

TABLE 1. Initial dry-weight water content (% , mean ± s.e.) of *Brassica tournefortii* seeds prior to treatment with karrikinolide

Treatment	Dry-weight seed water content
15 % RH	6.6 ± 0.5
47 % RH	10.1 ± 0.6
96 % RH	30.9 ± 0.2
Fully imbibed	86.3 ± 5.7
Re-dried (15 % RH)	7.0 ± 0.2

RH, relative humidity.

* Water content with mucilage was 319.7 ± 33.1 % d. wt.



RESULTS

KAR₁ sensitivity

For the seeds with no history of imbibition, seeds with lower seed water content were more sensitive to KAR₁, as more seeds germinated following shorter exposure times (Fig. 1). Seeds that were pre-equilibrated to 15 and 47% RH germinated to similar levels ($t = 1.73$, $P = 0.083$, d.f. = 4) and were the most sensitive to both 1 nM and 1 μM KAR₁; more than 50% of seeds germinated when exposed to 1 μM KAR₁ for as little as 3 min. In contrast, seeds from the 96% RH and fully imbibed treatments only germinated to levels above 50% when exposed to 1 μM KAR₁ for 1 h or longer. Across all treatments, the germination response to 1 nM KAR₁ was limited, yet for this concentration the effect of seed water content on KAR₁ sensitivity was still clear; only the 15 and 47% RH treatments germinated in response to 1 nM KAR₁, and only when exposed for longer than 1 h. All hydration treatments germinated to 100% following exposure to 1 μM KAR₁ for 14 d, confirming that the hydration treatments *per se* did not affect seed viability ($P \geq 0.837$, d.f. = 4).

A history of imbibition was found to influence the seed responses to KAR₁, as seeds that had been imbibed were less sensitive to KAR₁. Although the 15% RH seeds, which had never been imbibed, and the re-dried seeds had the same seed water content before exposure to the KAR₁ solutions (approx. 6.8% d. wt; Table 1), the re-dried seeds were less responsive to KAR₁ ($P < 0.001$, d.f. = 4). Indeed, the response of the re-dried seeds was indistinguishable from that of the fully imbibed seeds ($P = 0.189$, d.f. = 4), and only marginally different from the 96% RH seeds ($P = 0.049$, d.f. = 4), despite having a lower seed water content.

Seed imbibition

Seed water uptake was influenced by the initial hydration state of seeds. Seeds equilibrated at 15, 47 and 96% RH gained weight similarly, and were indistinguishable from the fully imbibed treatment within 3 min of imbibition ($P \geq 0.417$, d.f. = 4; Fig. 2). The re-dried seeds absorbed less water than the 15, 47 and 96% RH treatments in the first 45 min of imbibition ($P = 0.022$, d.f. = 1). All treatments reached their maximum seed weight, which was at least three times their dry-seed weight, within 3 h of imbibition. At 24 h, the re-dried seeds and fully imbibed seeds equilibrated at a lower weight than the other three treatments ($P < 0.001$, d.f. = 1), which had not previously imbibed, but all treatments lost weight between 3 and 24 h ($P = 0.01$, d.f. = 1).

Scanning electron microscopy

Scanning electron microscopy images showed that the mucilage layer of re-dried seeds was damaged relative to that of

FIG. 1. Germination of *Brassica tournefortii* seeds following exposure to 1 nM (closed circles) and 1 μM (open circles) KAR₁. Seeds were equilibrated to 15% RH, 47% RH, 96% RH, were fully imbibed, or were fully imbibed then dried to 15% ('re-dried') prior to exposure to KAR₁. Exposure durations were 0, 3 min, 1 h, 24 h or 14 d. Data are presented on a log scale as binomial estimates with asymmetrical error bars representing a 95% confidence interval ($n = 150$ seeds per data point).

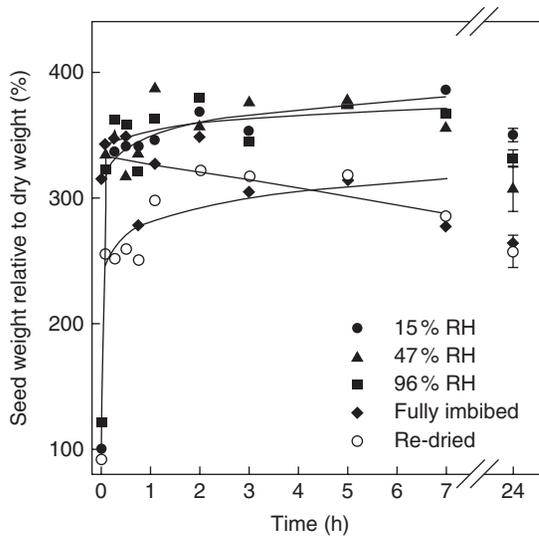


FIG. 2. Imbibition curves for *Brassica tournefortii* seeds pre-equilibrated to five different hydration states: 15% RH, 47% RH, 96% RH, fully imbibed, or fully imbibed then re-dried to 15% RH ('Re-dried'). For clarity, mean data ($n = 3$ replicates of 50 seeds) are presented with fitted exponential curves (except for fully imbibed, for which a linear curve was fitted) and ± 1 s.e. bars are only shown for the final time point.

seeds at 15% RH with no history of imbibition (Fig. 3). The mucilage layer visibly flaked away from the surface of re-dried seeds, and the morphology of the underlying testa of re-dried seeds appeared to be smoother.

Endogenous ABA, GA_3 and GA_4 concentrations

Seeds that had been imbibed contained approximately half the ABA concentration found in dry seeds with no history of imbibition (Fig. 4A), which was surprising given the imbibed seeds were more dormant. For the different hydration treatments without prior imbibition, ABA concentrations were found to be similar ($P \geq 0.765$, d.f. = 4); the fully imbibed and re-dried samples were found to have indistinguishable ABA concentrations ($P = 0.888$, d.f. = 4). Both GA_3 and GA_4 were present at low concentrations in all the seed batches, and were found not to differ significantly between the different hydration treatments (Fig. 4B, C; $P \geq 0.335$ and $P \geq 0.366$).

ABA, GA_3 and GA_4 sensitivity

Seeds with a history of being imbibed were more sensitive to exogenously applied ABA than seeds that had not been imbibed prior to exposure, but there were no differences in

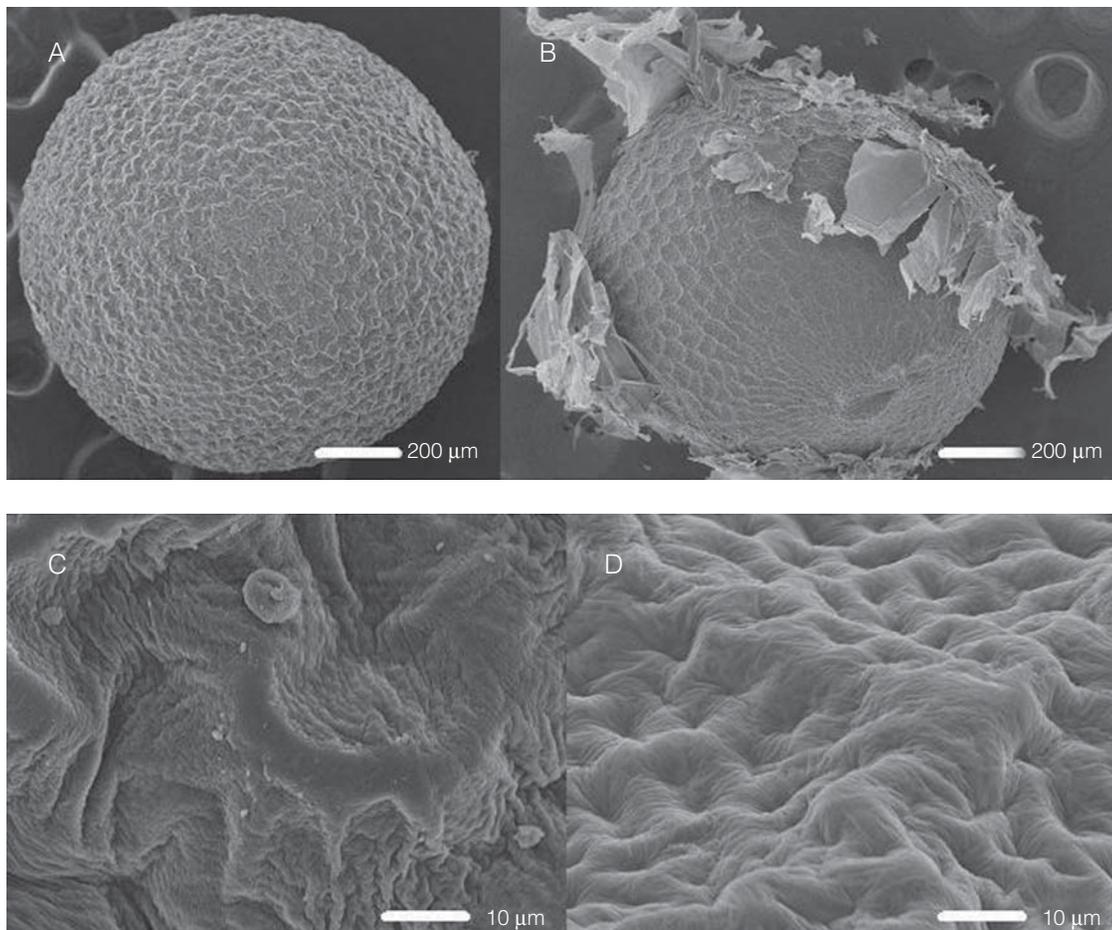


FIG. 3. Scanning electron microscope images of *Brassica tournefortii* seeds at 15% RH: seed with no history of imbibition (A, C) and seed that was re-dried following 18 h of imbibition (B, D).

sensitivity to either GA₃ or GA₄ (Fig. 5). Compared with the 15 % RH treatment, germination of the re-dried treatment was inhibited by 10 and 50 μM ABA ($P = 0.017$ and <0.001 ; d.f. = 2). Seeds in the fully imbibed treatment responded in the same way as the re-dried treatment ($P = 0.254$), but their germination was only distinguishable from the 15 % RH treatment at 50 μM ABA ($P < 0.001$). All hydration treatments responded equally to GA₃ and GA₄ ($P \geq 0.063$ and $P \geq 0.212$; d.f. = 2), with GA₄ stimulating germination at lower concentrations than GA₃.

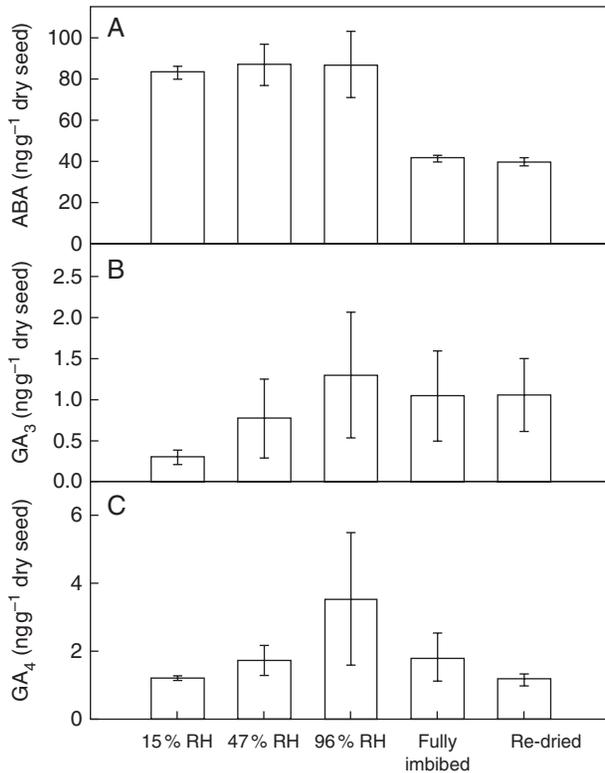


FIG. 4. The endogenous concentration (mean \pm 1 s.e., $n = 3$) of (A) ABA, (B) GA₃ and (C) GA₄ in *Brassica tournefortii* seeds pre-equilibrated to the five different hydration states: 15 % RH, 47 % RH, 96 % RH, fully imbibed, or fully imbibed then re-dried to 15 % RH ('Re-dried').

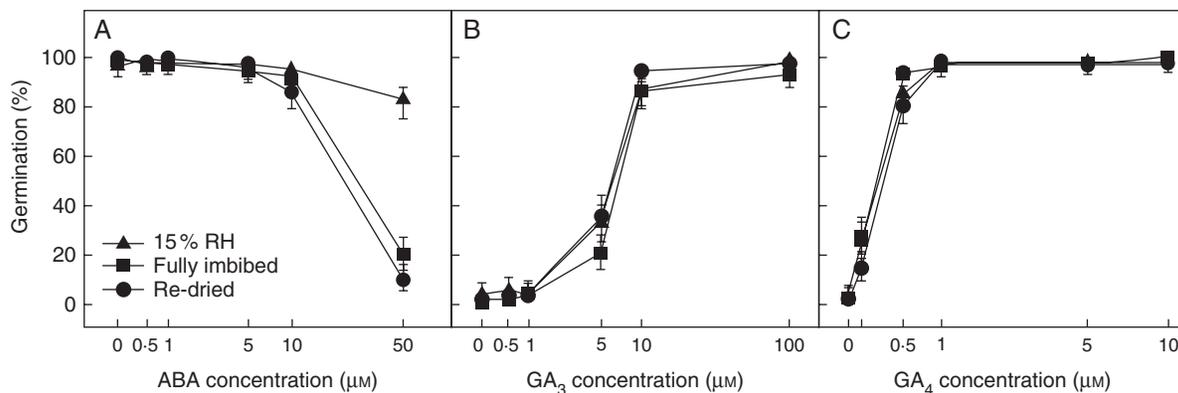


FIG. 5. Effect of exogenously applied ABA (A; ABA with 1 μM KAR₁), GA₃ (B) and GA₄ (C) on the germination of *Brassica tournefortii* seeds. Prior to plating on the relevant agar media, seeds were either dry with no history of imbibition ('15 % RH'), soaked in water for 18 h prior to treatment ('fully imbibed') or soaked for 18 h then re-dried to 15 % RH ('re-dried'). Data are presented as binomial estimates with asymmetrical error bars representing the 95 % confidence interval ($n = 150$ seeds per data point).

DISCUSSION

Our hypothesis that KAR₁ would stimulate more germination when applied to dry seeds than to seeds with higher water contents was accepted, but only for cases when the seeds had not previously imbibed water. Seeds with a history of imbibition were less sensitive to KAR₁, even when re-dried. Furthermore, re-dried seeds behaved similarly to fully imbibed seeds in germination assays at both concentrations of KAR₁, even though the fully imbibed seeds had higher water contents at the point of exposure to KAR₁. Accordingly, water content alone does not explain why the treatments responded differently to KAR₁. Thus, we conclude that both water content and the history of imbibition are important factors that influence the ability of KAR₁ to stimulate seed germination.

Our observations that the water content and history of imbibition of seeds influenced the sensitivity of *B. tournefortii* to KAR₁ may be explained by physical and physiological changes that occur when seeds imbibe. In a physical sense, the hydration state of seeds could influence how seeds respond to KAR₁ by dictating the nature of the forces driving the uptake of the stimulant in solution, and the quantity of solution absorbed. Additionally, the amount of water in seeds can determine their physiological activity by influencing the types and rates of reactions that occur (Walters *et al.*, 2005). In particular, when seeds imbibe they can synthesize, break down or change sensitivity to ABA and GA, which mediate germination (Grappin *et al.*, 2000; Ali-Rachedi *et al.*, 2004). Thus, we proposed three explanations for why hydration state influenced germination in response to KAR₁, namely that it affected (1) the physical uptake of the KAR₁ solution, (2) the endogenous concentration of ABA or sensitivity to ABA, and (3) the endogenous concentration of GA₃ and GA₄ or sensitivity to GA₃ and GA₄.

Imbibition curves for seeds of each hydration state afforded insight into the physical uptake of the KAR₁ solutions. The re-dried, 15, 47 and 96 % RH seeds at least doubled their weight in the first hour – probably due to having more negative matric potentials – whereas any uptake of the solution by the fully imbibed seeds was driven by osmosis. Yet the quantity and rate of solution absorption was not associated with

germination, as the re-dried seeds germinated like the fully imbibed seeds, despite having different water contents when initially exposed to KAR₁. The fully imbibed and re-dried seeds equilibrated at lower water contents relative to the other hydration treatments at 24 h, probably due to mucilage loss. This loss of mucilage also did not correlate with differences in germination as the fully imbibed and re-dried treatments had the same germination response as the 96 % RH treatment following 24 h of exposure to the KAR₁ solutions. Thus, differences in the volume of uptake and physical forces driving the uptake of solutions did not fully explain why the history of hydration had a significant effect on the germination response of seeds to KAR₁.

One possible physiological explanation for the results is that KAR₁ interacts with ABA concentration or ABA sensitivity, which change according to the hydration state of seeds (Finch-Savage and Leubner-Metzger, 2006). No link was found in the present study between endogenous concentrations of ABA and germination response, as ABA levels decreased with imbibition. Indeed, the lower concentration of ABA in the fully imbibed and re-dried treatments implies that dormancy is partially alleviated in these seeds (Ali-Rachedi et al., 2004; Feurtado et al., 2007), yet fewer of these seeds germinated with KAR₁. On the one hand, these results were surprising as we had reasoned that the poorer germination of previously imbibed seeds may be due to increased endogenous concentrations of ABA, which could override the stimulatory effect of KAR₁. On the other hand, the results are supported by other reports that ABA concentrations decline as seeds imbibe (Seo et al., 2006; Toh et al., 2008; Nelson et al., 2009), and are not always reflected in increased germination outcomes (Millar et al., 2006).

Despite the apparent lack of association between endogenous concentrations of ABA and the germination response of seeds, a different pattern with ABA sensitivity was observed. That is, seeds that had previously been imbibed were more sensitive to exogenous applications of ABA than were dry seeds. ABA sensitivity is moderated by a range of proteins that facilitate ABA signalling (Finkelstein et al., 2002), and it is plausible that transcription of genes for intermediates involved in signalling is favoured when seeds imbibe. Therefore, we reason that seeds become more sensitive to ABA during imbibition, and so become more receptive to what little ABA might be present in the seeds. In turn, the heightened sensitivity to ABA may obstruct the stimulatory effect of KAR₁, affording seeds a poorer germination response.

A second possible physiological explanation for the observed link between seed hydration state and germination in response to KAR₁ is that endogenous concentrations of gibberellins or the sensitivity of seeds to gibberellins are lower in imbibed seeds. Other studies have proposed interactions between gibberellins and smoke or KAR₁ signals (Schwachtje and Baldwin, 2004; Nelson et al., 2009). Schwachtje and Baldwin (2004) showed that exposure to smoke water enhanced the sensitivity of *Nicotiana attenuata* seeds to exogenous applications of GA₃, while Nelson et al. (2009) found no influence of KAR₁ on the sensitivity of *Arabidopsis thaliana* seeds to gibberellins. In the present study, there was no apparent connection between gibberellins and the germination response of seeds to KAR₁; neither the

endogenous concentrations of GA₃ and GA₄ nor the sensitivity of seeds to exogenous applications of GA₃ and GA₄ declined during imbibition or re-drying. Moreover, in contrast to the theory of Finch-Savage and Leubner-Metzger (2006) that high ABA/GA ratios pertain to greater dormancy and reduced germinability, the ratio of ABA to GA did not reflect the dormancy state and germinability of *B. tournefortii* seeds in this study. We conclude that ABA sensitivity is more important in determining seed germination characteristics than are the relative endogenous concentrations of ABA and gibberellins or the sensitivity to gibberellins. This finding is supported by Goggin et al. (2009) for *Lolium rigidum* seeds, who also identified ABA sensitivity as a key moderator of germinability.

The finding that higher seed water content and a history of imbibition can reduce the sensitivity of seeds to KAR₁ has clear implications for practice and research. First, we recommend that KAR₁ should be applied when seeds are dry to optimize the germination response. In a Mediterranean climate, where seeds experience an extended dry period following dispersal in late spring or early summer, KAR₁ may be most effective if applied prior to the onset of the late autumn rains, as was also observed for smoke applications in Roche et al. (1998). We recommend that future studies investigate how multiple hydration cycles, and hydration conditions that are common to other climates, such as cold- and warm-stratification, influence the receptivity of seeds to KAR₁. Finally, researchers should consider the hydration state and history of imbibition of seeds when conducting experiments using germination stimulants. As seen here, using only seeds in higher hydration states or those that had previously been imbibed could have led to the erroneous conclusion that the seed lot was not responsive to 1 nM KAR₁. It is plausible that germination with other commonly used stimulants such as smoke-water and nitrates may also depend on hydration state and history, especially given the observed effects of hydration state on ABA concentrations and sensitivity.

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